

APPLICATION HIGHLIGHT

Cardiomyocyte beating and cytotoxicity assays on the SpectraMax i3 Multi-Mode Microplate Platform with SpectraMax MiniMax Imaging Cytometer

Stem cell-derived human cardiomyocytes have phenotypic characteristics and electrophysiological profiles similar to those of native human cardiac cells. Cardiomyocytes in culture are able to form a beating syncytium, which behaves similarly to native cardiomyocytes. Oscillation of intracellular calcium levels occurring with synchronized contractions of the cells can be monitored using a calcium-sensitive dye, and treatment-induced changes in the pattern of oscillation can be monitored via changes in fluorescent signal over time.

Recently, it has been shown that changes in intracellular calcium flux associated with cardiomyocyte contractions can be monitored on a fluorescence imaging plate reader (FLIPR® Tetra System as well as SpectraMax® Paradigm System). Now, the SpectraMax® i3 Multi-Mode Microplate Platform with SpectraMax® MiniMax™ Imaging Cytometer from Molecular Devices enables detection of cardiomyocyte beating and fluorescence imaging of cell viability in a single instrument. Compound-induced changes in observed beating rate can be correlated to cell viability for a more accurate assessment of cardiotoxicity.

A cardiac beating assay was performed on the SpectraMax i3 Multi-Mode Microplate Platform using the EarlyTox™ Cardiotoxicity Kit from Molecular Devices® for assessment of compound effects upon cardiomyocytes. iCell Cardiomyocytes, human induced pluripotent stem cell (iPSC)-derived cardiomyocytes from Cellular Dynamics International, were cultured in 384-well microplates until a synchronously beating layer formed. Cells were then treated with compounds from

the Screen-Well Cardiotoxicity Library (Enzo Life Sciences) at 10 μ M for 24 hours and assessed for compound effects on beating and viability.

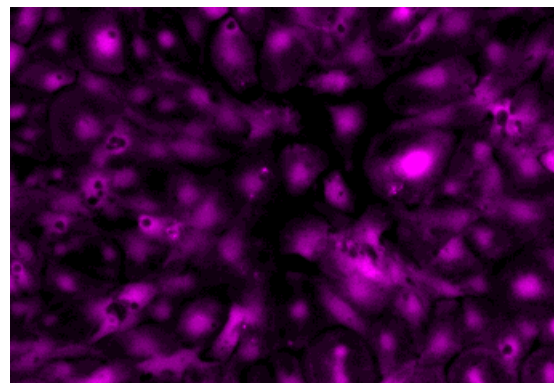
Cardiomyocyte beating parameters were measured on the SpectraMax i3 Platform using a kinetic, well-by-well fluorescence read mode and analyzed using Peak Pro software algorithms in SoftMax® Pro Software. Sixteen analysis parameters, including peak count, peak frequency, peak amplitude, peak width, decay time, and several others, were calculated. In this example peak count, the number of peaks measured during a 25-second reading, was used to quantify the effects of compounds on cardiac beating.

To obtain additional information on possible toxicity of compounds that caused a decrease in the peak count, live cells were stained with the green-fluorescent viability dye calcein AM (Life Technologies) after compound treatment and beat measurement. Cells were imaged using the SpectraMax MiniMax Imaging Cytometer, a field-upgradable option for the SpectraMax i3 Platform. Several cell analysis methods are available in SoftMax Pro Software, including cell count, cell proliferation, and marker expression. Changes in cell viability due to compound toxicity were analyzed using the cell proliferation analysis protocol to measure the area covered by live, calcein AM-stained cells in each well.

Using the combined analysis of cardiomyocyte beating and viability imaging, compounds with differing effects on cells were identified. The average peak count for control wells was 12 peaks per 25 second read time, and wells in which peak

Benefits

- Obtain more biologically relevant data by combining microplate reader assays with imaging cytometry.
- No imaging expertise required with intuitive SoftMax Pro Software user interface
- The only multi-mode platform that evolves to meet future application needs with user-installable application cartridges



count was significantly reduced could be easily distinguished by the software.

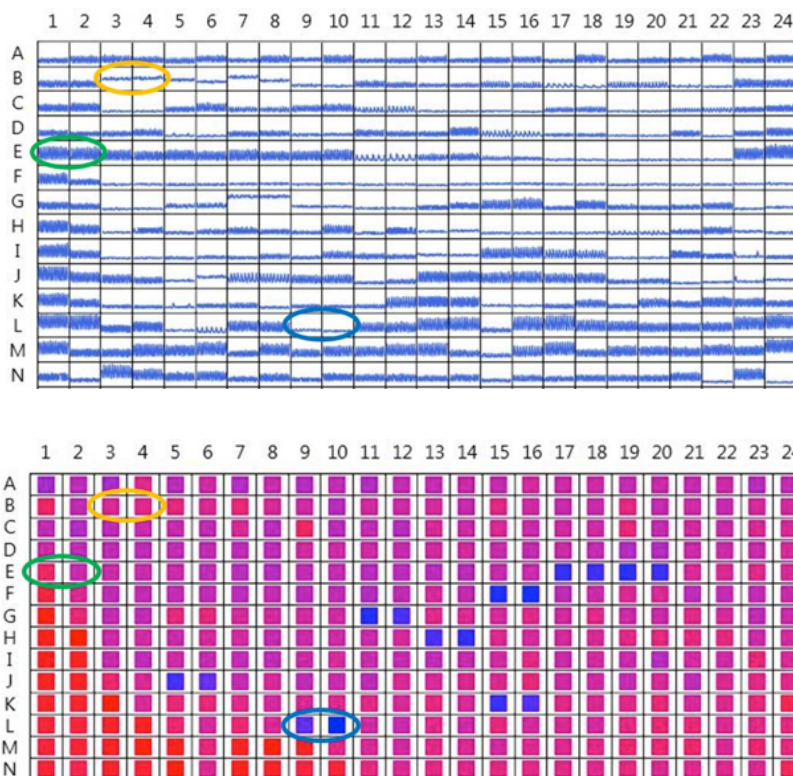
Some compounds were identified as generally cytotoxic, causing a significant reduction in peak count and a decrease in percent area covered by viable cells. Other compounds adversely affected cardiomyocyte beating without impacting cell viability, as evidenced by a low peak count and a high percentage of area covered by viable cells. Figure 1 shows plate views of the data for both cardiac beating (top) and area covered by viable cells (bottom).

Plate views offer an easy visual check for correlation between the effects of compounds on beating patterns and cell viability. In Figure 2, more detailed views of individual kinetic traces and representative images are shown for compounds with different effects on cardiomyocytes.

Several compounds that dramatically reduced the peak count were shown to have little or no effect on overall cell viability. For example, digoxin, a drug widely used in the treatment of arrhythmias, reduced peak count from the roughly 12 beats per 25-second reading down to 1 or 2 beats. However, cell covered area was 71% compared to 82% in DMSO-treated controls—not a significant reduction in cell viability. The dopamine inverse agonist haloperidol had a distinct effect on beating profile without impacting viability. Other compounds, like staurosporine, greatly reduced both cell viability and peak count (Figure 2).

Based on results from the compound library screen, compounds with observed effects on beating rate and cytotoxicity were selected for more detailed study. Representative compounds were added to cardiomyocytes in a 1:3 dilution series to determine IC₅₀ values. As before, cardiomyocyte contractions were visualized using the EarlyTox Cardiotoxicity dye, and beating profiles measured on the SpectraMax i3 Platform.

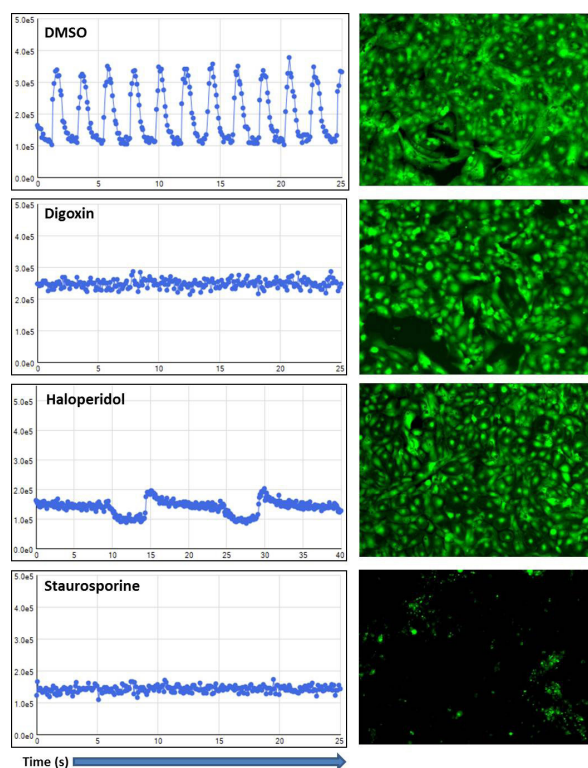
Figure 1. Multi-well views of cardiac beating and viability data



Top: Representative kinetic traces from cardiomyocytes treated with compounds from the ENZO Cardiotoxicity Library. Bottom: Heat map of cell viability imaging results, expressed as percentage of the well covered by live cells, for the same assay plate as above. Blue-shaded wells indicate low viability, while red-shaded wells indicate highest viability. Examples of duplicate DMSO-treated control wells are circled in green on both plate views. Digoxin-treated wells showing reduction in peak count without loss of viability are circled in yellow, and staurosporine-treated wells where both peak count and viability are significantly reduced are circled in blue.

Figure 2. Comparing peak data to cell viability

Detailed views of kinetic traces and images of cells treated with DMSO (control), digoxin, haloperidol, or staurosporine. Digoxin and haloperidol decreased the number of peaks measured in beating cells but had no significant effect on viability, while staurosporine negatively affected both beating and viability.



Data were analyzed using Peak Pro algorithms in the SoftMax Pro Software (Figure 3, Top). After the beating was measured, the same cells were stained with calcein AM and imaged using the MiniMax Imaging Cytometer to determine cell viability. Concentration-response curves were plotted as area covered vs. compound concentration (Figure 3, Bottom).

Conclusions

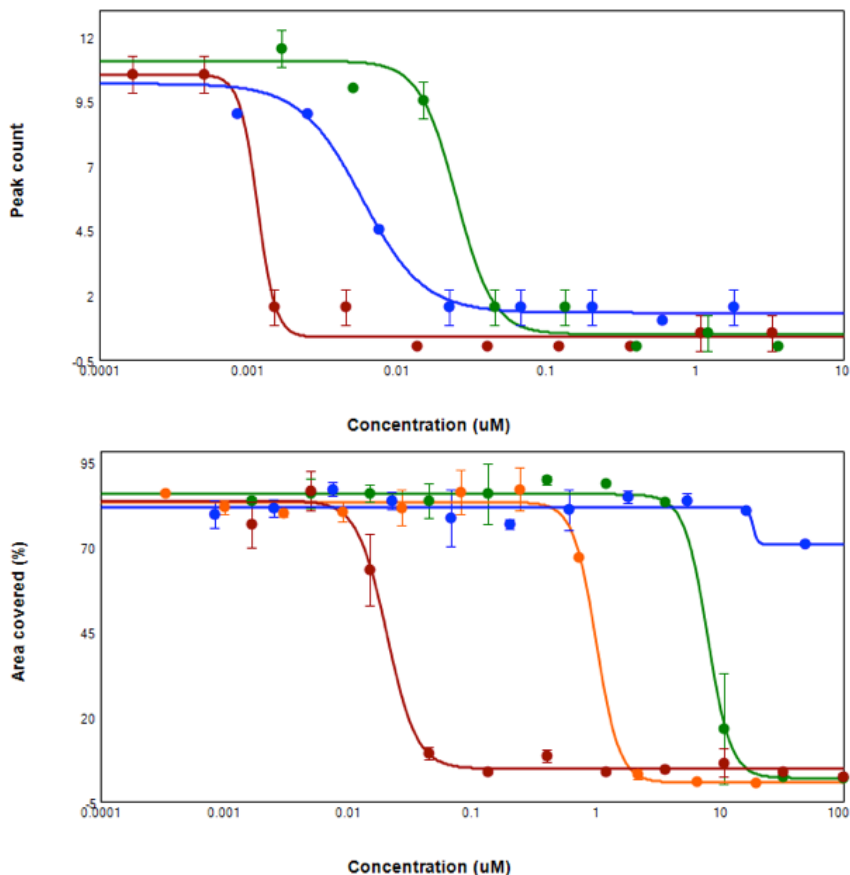
The SpectraMax i3 Platform with SpectraMax MiniMax Imaging Cytometer enables detection of cardiomyocyte beating and fluorescence imaging of cell viability in a single instrument, with integrated data analysis provided by SoftMax Pro Software. Compound-induced changes in observed beating rate, or abnormal rhythms, can be correlated to cell viability. Concentration-dependent effects on cardiomyocyte beating on the SpectraMax i3 Platform for several reference compounds have been demonstrated.

other means than apoptosis. Increased cell death was observed with heat shock, particularly after 24 hours. Stressed cells had a higher overall percentage of dead cells than did the normal population. These results match reasonably well with the apoptosis result, suggesting that all or most of the cell death observed is due to apoptosis.

The MiniMax Imaging Cytometer was used to image cytotoxic effects in the same wells that were monitored for cardiomyocyte beating. Measuring functional readouts, like calcium flux and cell viability, in the very same wells, provides complementary information on the impact of compounds upon cardiomyocyte function and overall cell toxicity.

The SpectraMax i3 Platform with MiniMax Imaging Cytometer seamlessly combines the benefits of imaging with the versatility of multi-mode plate reader assays. SoftMax

Figure 3. Concentration-response curves for peak count and cell viability



Top: Concentration-response curves based on peak counts for cells treated with valinomycin (red), digoxin (blue), and amiodarone (green). Bottom: Concentration-response curves for viability of cells treated with valinomycin (red), digoxin (blue), amiodarone (green), and staurosporine (orange).

Pro Software offers a simple workflow from data collection to analysis, and user-upgradeable cartridge options make this system an intelligent investment. Learn more about the system: www.moleculardevices.com/SpectraMax This article originally appeared in the September 1, 2013 issue of GEN (Genetic Engineering & Biotechnology News).

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