

APPLICATION NOTE

Complete cGMP workflow solution using the CatchPoint cGMP Fluorescent Assay Kit

Introduction

In this study, we demonstrate how the CatchPoint® cGMP Fluorescent Assay Kit combined with the SpectraMax® i3 Multi-Mode Detection Platform can be used to measure cGMP in RFL-6 cells treated with atrial natriuretic peptide (ANP) [1, 2]. See assay workflow in Figure 1.

The second messenger 3', 5' cyclic guanosine monophosphate (cGMP) and its associated proteins have been shown to be integral in regulating physiological responses such as smooth muscle relaxation, vasodilation, kidney function, phototransduction, and penile erection [3–7].

ANP, a hormone found in many vertebrates, signals through cGMP to reduce blood pressure by stimulating the excretion of sodium and water from the kidneys and relaxing vascular smooth muscle [8, 9]. Binding of ANP to its receptor leads to conversion of GTP to cGMP, which in turn activates protein kinase G, causing activation of a number of biologically relevant targets.

The CatchPoint cGMP Fluorescent Assay Kit measures cGMP levels via a competitive immunoassay (Figure 2). The assay requires only a single washing step, and readings can be taken in as little as 10 minutes or as long as 24 hours following substrate addition.

Materials

- CatchPoint cGMP Fluorescent Assay Kit (Molecular Devices cat. #R8074)
- RFL-6 rat lung fibroblast cells (ATCC cat. #CCL-192)
- F-12K Medium (Kaighn's Modification of Ham's F-12 Medium, ATCC cat. #30-2004)
 - Fetal bovine serum (Gemini Bio-Products cat. #100-106)
 - Penicillin-streptomycin (Life Technologies cat. #15070-063)
- Krebs-Ringer bicarbonate buffer (KRGB buffer, Sigma cat. #K4002)
 - Sodium bicarbonate (Sigma, cat. #S5761)
- Bovine serum albumin (Sigma cat. # 9048-46-8)
- Phosphate-buffered saline (PBS, Life Technologies cat. #10010)
- Human ANP (Sigma cat. #A-1663)
- Phosphodiesterase inhibitor, 3-isobutyl-1methylxanthine (IBMX, Sigma cat. #I7018)

Benefits

- Accurate measurements of cGMP in cell-based assays
- Assay requires only a single wash step
- High signal stability (10 minutes to 24 hours)
- Z' factor of 0.82

- 3% hydrogen peroxide (H₂O₂) solution
- 96-well, clear-bottom, tissue culturetreated microplates (Corning cat. #3603)
- SpectraMax i3 Multi-Mode Microplate Reader
- SpectraMax® MiniMax™ 300 Imaging Cvtometer

Confluence **Fluorescence Cell preparation** CatchPoint assay Plate wash Data analysis measurement measurement RFL-6 cells SpectraMax CatchPoint cGMP MultiWash+ SpectraMax i3 SoftMax Pro MiniMax 300 Fluorescent Microplate Washer Multi-Mode Software Imaging Cytometer Microplate Reader Assay Kit

Figure 1: CatchPoint cGMP workflow.

Methods

RFL-6 cells were cultured in complete culture media (F-12K medium + 20% FBS + 1% pen/strep) to at least 90% confluence. Cells were harvested with 0.05% trypsin and then seeded into a tissue culture-treated, 96-well, black-wall, clear-bottom microplate at 20,000 cells/well (15,000-50,000 cells/well is an acceptable range). Cells were allowed to adhere for at least 18 hours in a 37°C incubator with 5% CO₂.

On the day of assay, cell confluence was measured using the transmitted light channel of the SpectraMax MiniMax 300 Imaging Cytometer and StainFree™ Cell Detection Technology, which enables accurate measurement of cell counts or percent area covered by cells (confluence) for quality control purposes. Rows of cells with high cell confluence and low inter-well variability were selected for the CatchPoint assay (Figure 3). Cells were incubated in pre-stimulation buffer, KRBG buffer containing 0.75 mM IBMX, for 10 minutes at room temperature. Cells were then treated with ANP starting at 1 µM with a 1:3 dilution series for 15 minutes at 37°C. Following treatment, cells were lysed according to the CatchPoint assay protocol.

The CatchPoint assay was performed as described in the product insert. A cGMP calibrator curve was performed to verify assay performance, as well as provide a means of calculating the amount of cGMP in cellular assay samples. Samples were read on the SpectraMax i3 Multi-Mode Microplate Reader 60 minutes after StopLight Red substrate addition. All data analysis and curve fitting were performed with SoftMax® Pro Software. A preconfigured protocol is available in the software's protocol library.

Results

To assess cell confluence before cell pre-stimulation and treatment, the MiniMax cytometer and StainFree technology were used to compare percent cell confluence among different rows of the microplate (Figure 3). Row B was chosen for assay due to consistent percent confluence from well to well. Triplicate samples were taken from each treated well and assayed.

Figures 4 and 5 show results for the CatchPoint cGMP calibrator curve and ANP response cell-based assay. Both curves

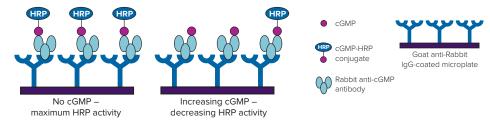


Figure 2: CatchPoint cGMP assay mechanism. Unlabeled cGMP produced by cells competes with cGMP-HRP conjugate for binding to anti-cGMP antibodies. Increasing amounts of cellular cGMP are detected as decreasing HRP activity.

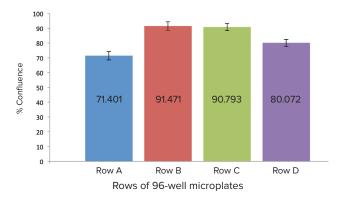


Figure 3: Relative cell confluence of RFL-6 cell-seeded wells. Row B was chosen for the assay because of high-well confluence and low inter-well variability.

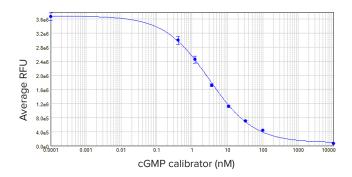


Figure 4: cGMP calibrator curve. EC₅₀ was 3.1 nM, similar to previously published results. Samples were run in triplicate.

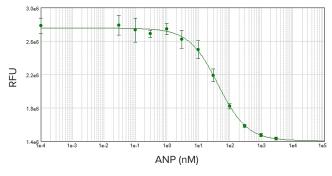


Figure 5: ANP concentration-response curve starting at 1000 nM with 3-fold dilution series. EC_{50} was 41 nM, consistent with previously published data. Samples were run in duplicate.

were generated using the 4-parameter curve fit in SoftMax Pro Software. The calibrator curve had an EC $_{50}$ of 3.1 nM, which agreed closely with previously published data. The Z' factor was 0.82.

The ANP concentration-response observed in RFL-6 cells had an EC $_{50}$ of 41 nM, which was also consistent with previously reported results.

Conclusion

The CatchPoint Cyclic GMP Fluorescent Assay Kit is accurate and robust for measuring cGMP in cell-based assays. The assay's high signal stability (can be read 10 minutes to 24 hours after substrate addition), excellent Z' factor, and ability to be performed on plate readers with a fluorescence intensity detection mode make the assay a versatile choice for high-throughput screening. The SpectraMax MiniMax 300 Imaging Cytometer and StainFree analysis offer an additional level of quality control for cell-based assays.

Ordering information		
Reagent	Description	Part number
CatchPoint cGMP 96-well Explorer Kit	192 reactions	R8074
CatchPoint cGMP 96-well Bulk Kit	960 reactions	R8075
CatchPoint cGMP 384-well Explorer Kit	768 reactions	R8065
CatchPoint cGMP 384-well Bulk Kit	7680 reactions	R8066

Compatible with these Molecular Devices systems



SpectraMax i3 Platform with SpectraMax MiniMax 300 Imaging Cytometer



MultiWash+ Microplate Washer

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- Leitman, Dale C., et al. "Atrial natriuretic peptide binding, cross-linking, and stimulation of cyclic GMP accumulation and particulate guanylate cyclase activity in cultured cells." *Journal of Biological Chemistry* 263.8 (1988): 3720-3728.

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